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# Title: Rapid screening of COVID-19 patients by White blood cells scattergrams, a study on 381 patients

Revisions are marked in bold red.

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**Running title**: Rapid screening of COVID-19 patients by WBC scattergrams

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#### Abstract

Complementary tools are warranted to increase the sensitivity of the initial testing for COVID-19. We identified a specific "sandglass" aspect on the white blood cells scattergram of COVID-19 patients reflecting the presence of circulating plasmacytoid lymphocytes. Patients were dichotomized as COVID-19 positive or negative based on RT-PCR and chest CT scan results. Sensitivity and specificity of the "sandglass" aspect were **85.9%** and **83.5%** respectively. The positive predictive value was **94.3%**.

Our findings provide a non-invasive and simple tool to quickly categorize symptomatic patients as either COVID-19 probable or improbable especially when RT-PCR and/or chest CT are not rapidly available.

#### INTRODUCTION

The novel coronavirus SARS-CoV-2, responsible for COVID-19 confronts the health community with major challenges.(1) Early diagnosis of COVID-19 is crucial for the optimal management of infected patients to control viral spread. The standard test for COVID-19 remains the Reverse Transcriptase Polymerase Chain Reaction (RT-PCR) to detect viral RNA from clinical samples. RT-PCR is specific but **lacks sensitivity**.(2–4) Complementary tools are warranted to increase the sensitivity of the initial testing of COVID-19 patients.

Complete blood count (CBC) is a routine test during initial biological assessment of patients. CBC analyzers such as SYSMEX® (Japan), provide a white blood cells (WBC) differential fluorescence (WDF) scattergram, displaying a classification of WBC based on their morphology and their intracellular components. Each type of leucocyte is always displayed on the same area. The different clusters of leucocytes displayed on the WDF match with the visual examination by optical microscopy.

During this outbreak, we have noticed a recurrent atypical aspect on the WDF of COVID-19 patients. We therefore decided to evaluate the sensitivity and specificity of our finding in order to propose WDF as a screening tool for COVID-19.

## **METHODS**

Patients admitted at Versailles Hospital suspected of having COVID-19 were eligible if symptoms were present for 3 or more days and if RT-PCR and a chest CT were performed (Supplementary Figure 1).

CBC was performed using an XN3100 analyzer (SYSMEX®). WDF were assessed blindly by two readers. Presence of the new pattern was considered WDF positive (WDF+), all other patterns were considered negative (WDF-). Blood cell morphology was assessed by microscopy (Figure 1 and Supplementary Figure 2).

RNA was extracted from clinical samples obtained via upper or lower respiratory tract swabs or aspirates. RT-PCR assays were performed on APPLIED BIOSYSTEMS® analyzers (USA), following the National Reference Center protocol (Pasteur Institute). Results were concluded as positive (RT-PCR<sup>+</sup>) if amplification of SARS-CoV2 cDNA was observed after 40 cycles.

Chest CT **scans** were performed on GENERAL ELECTRIC® scanners (USA) and classified as typical (CT+) or not (CT-) for COVID-19 according to published definition.(3,5-7)

"Index test" was the WDF pattern on the CBC performed at admission time, whereas the "reference test" was a diagnostic algorithm combining RT-PCR and CT results, as recommended by recent studies.(3,5) **We excluded patients with symptoms for less than 3 days to overcome the "grey-zone" of the chest CT.** Patients with at least one RT-PCR<sup>+</sup> and/or CT<sup>+</sup> were considered as COVID-19 positive (COVID-19<sup>+</sup>) whereas patients with RT-PCR<sup>-</sup> and CT<sup>-</sup> were considered as COVID-19 negative (COVID-19<sup>-</sup>). WDF and Chest CT interpretations were blinded.

Once dichotomized (COVID-19<sup>+/-</sup>), diagnostic performances of WDF were calculated.

All statistical analyses were performed using R version 3.6.1 (R Core Team 2019). Patient's baseline characteristics were compared by non-parametric tests, either the exact Fisher's test for qualitative variables or the Kruskal-Wallis test for quantitative variables.

This study was conducted in accordance with the French CNIL (commission informatique et libertés) regulations.

## RESULTS

We noticed a recurrent atypical aspect on the WDF scattergram of COVID-19 patients. This aspect, named the "sandglass" pattern, consisted **of** a discontinuous cluster of lymphocytes characterized by the presence of more than 4 dots in the upper **graduation** of the scattergram, where plasmacytoid lymphocytes are usually plotted(8,9). This observation was reinforced by the presence of circulating plasmacytoid lymphocytes on blood smears from patients with COVID-19 whereas large hyperbasophilic lymphocytes, normally seen in other viral infections, were absent (Figure 1). The 4 dots threshold was derived from the receiver operating characteristic (ROC) curve to maximize the weighted Youden index(10) (Supplementary Figure 3).

We then retrospectively analyzed 381 WDF from symptomatic adults admitted at Versailles Hospital from March 16<sup>th</sup> to April 5<sup>th</sup> 2020 (Median age: 61 years [18-99], sex ratio M/F: 1.47). Complete characteristics of patients are reported in Table 1.

In summary, 57% (216/381) of the patients were hospitalized including 36 patients (9%) immediately admitted to the intensive care unit for an acute respiratory distress syndrome. Loss of

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smell/taste (33/290) and lymphopenia (159/290) were largely reported in COVID-19<sup>+</sup> patients versus COVID-19<sup>-</sup>.(7,11–13)

The COVID-19 status confirmation was available within 1 day for 353/381 (93%) patients (range: 0-3 days). Of the 381 patients studied 290 (76%) were COVID-19<sup>+</sup> and 91 (24%) were COVID-19<sup>-</sup>. Among COVID-19<sup>+</sup> patients, 247 (85%) had RT-PCR<sup>+</sup>/CT<sup>+</sup>, 35 (12%) had RT-PCR<sup>-</sup>/CT<sup>+</sup> and 8 (3%) had RT-PCR<sup>+</sup>/CT<sup>-</sup>.

Interestingly, 25 COVID-19<sup>+</sup> patients with WDF<sup>-</sup> had a further CBC available, and the WDF became positive for 19 (76%) patients within 1-2 days. For the 15 COVID-19<sup>-</sup> patients with WDF<sup>+</sup>, a diagnosis of clinically documented pneumonia (10/15) or dyspnea (3/15), flu-like syndrome (1/15), or vaso-occlusive crisis (1/15) was finally made.

Using COVID-19<sup>+</sup> group as reference, we validated the performance of the WDF "sandglass" pattern as a screening tool for COVID-19. ROC curve was plotted and showed good discriminative performances of WDF with an area under the curve of 0.870 [95%CI: 0.830-0.910] (Supplementary Figure 3). Using 4-dots threshold, the diagnostic performances were: sensitivity: 85.9% [CI: 81.3-89.7], specificity: 83.5% [95%CI: 74.3-90.5] positive predictive value (PPV): 94.3% [95%CI: 90.8-96.8], negative predictive value (NPV): 65.0% [95%CI: 55.6-73.5], positive likelihood ratio: 5.2 [95%CI: 3.3-8.3] and negative likelihood ratio: 0.17 [95%CI: 0.13-0.23].

We then applied our test to a validation cohort of 170 WDF from patients infected with a well-defined pathogen (85 SARS-CoV-2, 54 Influenza virus, 19 Epstein-Barr virus, 8 *Mycoplasma pneumoniae*, and 4 Parvovirus B19) and found a sensitivity to distinguish COVID-19 versus other infections of 88.2% [95%CI: 79.4-94.2] and a specificity of 83.5% [95%CI: 73.9-90.7].

#### **DISCUSSION**

We report here a specific and original "sandglass" aspect on the WDF scattergram of COVID-19 patients. We hypothesize that this pattern reflects the presence of circulating plasmacytoid lymphocytes as observed from our careful blood smears examination of COVID-19 patients(8,9). Circulating plasmacytoid lymphocytes, absent in healthy people, have previously been **reported** in COVID-19(14,15) and deserve further immunological explorations. We showed that WDF is a highly reliable screening test to detect COVID-19 patients with 85.9% sensitivity and 83.5% specificity. It remains a simple, rapid, inexpensive, and non-invasive method. Due to COVID-19 associated lymphopenia,(7,11,12) WDF analysis appears more accurate than blood smear examination. If confirmed, detection of circulating plasmacytoid lymphocytes can be a useful alternative for centers where WDF is not available.

Our study however presents some limitations: First, it is a monocentric study carried out using a specified type of CBC analyzers. However, SYSMEX® analyzers are largely available in clinical institutions all over the world. This report may allow other laboratories and hospitals to confirm our results and provide multicentric data. Second, in order to exclude undetermined cases and reduce potentially wrong dichotomization resulting from early negative CT,(5–7) we excluded early symptomatic patients. Thus, prevalence of COVID-19 cases was higher than in the general population for which RT-PCR was required, and therefore PPV may be overestimated while NPV underestimated.

Based on this retrospective study, we conclude that WDF analysis can be implemented during the SARS-CoV-2 pandemic to quickly categorize symptomatic patients as either COVID-19 probable or improbable, depending **on** the presence of the plasmacytoid lymphocytes cluster on their scattergram.

Finally, given that CBC is available within few minutes, the "sandglass" WDF pattern may be a valuable tool assisting clinicians to pilot the medical management of symptomatic patients suspected of having COVID-19 at time of admission in hospitals.

This simple tool may be of particular importance (i) when RT-PCR and/or chest CT are not rapidly available, (ii) to decide to repeat the RT-PCR, (iii) in addition to other diagnostic tools such as chest CT and (iv) for patients for whom the diagnosis was not initially suspected.

We are now conducting a prospective validation cohort to derive a new algorithm combining RT-PCR, chest CT and WDF in order to facilitate the initial management of symptomatic patients suspected of having COVID-19.

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**Authorship contributions:** J.O. conceptualized, designed the study and wrote the first draft. J.O., M.T., F.D., R.F., C.F., D.B. and V.R analyzed data. J.L. provided statistical analysis. All authors provided critical revision of the manuscript.

Conflict of Interest Disclosures: The authors declare no competing financial interests

**Data sharing:** All data and materials used in this work are available based on reasonable request to the corresponding author.

### REFERENCES

5.

12.

- WHO/Europe | Coronavirus disease (COVID-19) outbreak [Internet]. [cited 2020 Apr 23]. Available from: http://www.euro.who.int/en/health-topics/health-emergencies/coronavirus-covid-19
- Ai T, Yang Z, Hou H, Zhan C, Chen C, Lv W, et al. Correlation of Chest CT and RT-PCR Testing in Coronavirus Disease 2019 (COVID-19) in China: A Report of 1014 Cases. Radiology. 2020 Feb 26;200642.
- Kim H, Hong H, Yoon SH. Diagnostic Performance of CT and Reverse Transcriptase-Polymerase Chain Reaction for Coronavirus Disease 2019: A Meta-Analysis. Radiology. 2020 Apr 17;201343.
- 4. Yang Y, Yang M, Shen C, Wang F, Yuan J, Li J, et al. Evaluating the accuracy of different respiratory specimens in the laboratory diagnosis and monitoring the viral shedding of 2019-nCoV infections. medRxiv. 2020 Feb 17;2020.02.11.20021493.
- 5. Shi H, Han X, Jiang N, Cao Y, Alwalid O, Gu J, et al. Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. Lancet Infect Dis. 2020;20(4):425–34.
- 6. Bai HX, Hsieh B, Xiong Z, Halsey K, Choi JW, Tran TML, et al. Performance of radiologists in differentiating COVID-19 from viral pneumonia on chest CT. Radiology. 2020 Mar 10;200823.
  - Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. JAMA. 2020 Feb 7;
    - van Mirre E, Vrielink GJ, Tjon-a-Tsoi N, Hendriks H, de Kieviet W, ten Boekel E. Sensitivity and specificity of the high fluorescent lymphocyte count-gate on the Sysmex XE-5000 hematology analyzer for detection of peripheral plasma cells. Clin Chem Lab Med. 2011 Apr;49(4):685–8.
    - Linssen J, Jennissen V, Hildmann J, Reisinger E, Schindler J, Malchau G, et al. Identification and quantification of high fluorescence-stained lymphocytes as antibody synthesizing/secreting cells using the automated routine hematology analyzer XE-2100. Cytometry B Clin Cytom. 2007 May;72(3):157–66.
  - Perkins NJ, Schisterman EF. The inconsistency of "optimal" cutpoints obtained using two criteria based on the receiver operating characteristic curve. Am J Epidemiol. 2006 Apr 1;163(7):670–5.
  - Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet Lond Engl. 2020 15;395(10223):507–13.
  - Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet Lond Engl. 2020 15;395(10223):497–506.

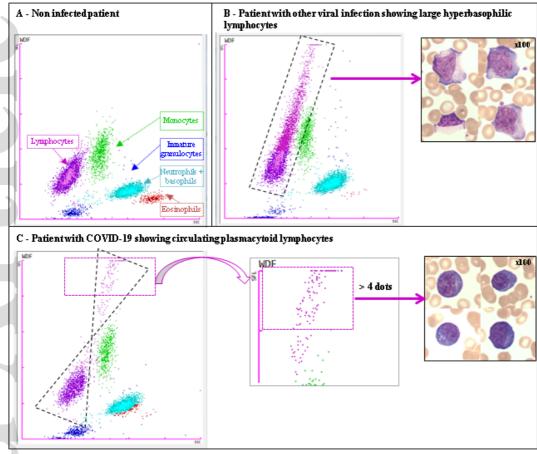
- 14. 15.
- 13. Giacomelli A, Pezzati L, Conti F, Bernacchia D, Siano M, Oreni L, et al. Self-reported Olfactory and Taste
  Disorders in Patients With Severe Acute Respiratory Coronavirus 2 Infection: A Cross-sectional Study. Clin
  Infect Dis [Internet]. [cited 2020 Apr 20]; Available from:
  https://academic.oup.com/cid/article/doi/10.1093/cid/ciaa330/5811989
  - 14. Foldes D, Hinton R, Arami S, Bain BJ. Plasmacytoid lymphocytes in SARS-CoV-2 infection (Covid-19). Am J Hematol. 2020 Apr 16;
    - Chong VCL, Lim KGE, Fan BE, Chan SSW, Ong KH, Kuperan P. Reactive lymphocytes in patients with Covid-19. Br J Haematol [Internet]. [cited 2020 May 18];n/a(n/a). Available from: https://onlinelibrary.wiley.com/doi/abs/10.1111/bjh.16690

## Tables and Figure legend

Figure 1. Atypical white blood cells scattergram on patients suspected of having COVID-19 and matching aspects on the blood smear.

**A** – Example of a normal WDF scattergram from healthy patient (no or less than 4 dots on the upper **graduation**). After permeabilization of the leucocyte membrane and intracellular staining, the WDF scattergram (White blood cells (WBC) Differential Fluorescence, XN3100 SYSMEX) can differentiate WBC depending on their morphology (side scattered light, SSC, x-axis) and the content of RNA/DNA (side fluorescent light, SFL, y-axis). Each dot represents one analyzed cell. Each type of leucocyte is always displayed on the same area. The different clusters of leucocytes displayed on the WDF match with the visual examination by optical microscopy (May-Grunewald Giemsa staining, original magnification x100).

- **B** Example of a WDF scattergram usually observed in case of other viral infections. This aspect consisted of a continuous cluster of lymphocytes and large hyperbasophilic lymphocytes as observed on the blood smear.
- C Example of an atypical aspect on the WDF scattergram of patients having COVID-19. This aspect consisted of a discontinuous cluster of lymphocytes characterized by the presence of more than 4 dots in the upper **graduation** of the scattergram ("sandglass" aspect), where plasmacytoid lymphocytes are usually plotted. This pattern reflects the presence of circulating plasmacytoid lymphocytes as observed from a careful analysis of blood smears from COVID-19 patients.



**Table 1. Characteristics of the cohort.** 

	OVERALL N=381	COVID-19 <sup>+</sup>	COVID-19- N=91	P-Values
		N=290		
DEMOGRAPHICS				
Median of age / [Range]	61 [18-99]	62 [21-99]	57 [18-94]	0.33
(years)				
<50 years, No. (%)	102 (27%)	66 (23%)	36 (40%)	
≥ 70 years, No. (%)	131 (34%)	100 (35%)	31 (34%)	
Male (No.) / Female (No.)	227 / 154	185 / 105	42 / 49	0.003
CLINICAL FEATURES (NS	= 2)			
SYMPTOMS FREQUENTLY	Y OBSERVED <sup>7,9</sup> -	11		
Fever	296 (78%)	243 (84%)	53 (58%)	< 0.001
Cough	246 (65%)	194 (67%)	52 (57%)	0.10
Dyspnea	240 (63%)	186 (64%)	54 (59%)	0.46
ARDS	37 (10%)	31 (11%)	6 (7%)	0.31
Loss of smell or taste	36 (9%)	33(11%)	3 (3%)	0.023
Confusion	11 (3%)	8 (3%)	3 (3%)	0.79
Headache	50 (13%)	39 (13%)	11 (12%)	0.86
Chest pain	42 (11%)	24 (8%)	18 (20%)	0.004
Asthenia	150 (39%)	128 (44%)	22 (24%)	< 0.001
Flu-like syndrome	103 (27%)	86 (30%)	17 (19%)	0.043
Digestive disorders	79 (21%)	62 (21%)	17 (19%)	0.66
Duration of symptoms at adn	nission time (days	) <sup>b</sup>		
Mean [range]	7.3 [3-30]	7.7 [3-30]	6.2 [3-30]	
Median	7	7	3	
BECOMING				
Non hospitalized	42 (11%)	19 (7%)	23 (25%)	
Pre-COVID Unita	121 (32%)	88 (30%)	33 (36%)	
Hospitalized	216 (57%)	182 (63%)	34 (37%)	
Among ICU	36 (9%)	30 (10%)	6 (7%)	
BIOLOGICAL FEATURES				
Median time interval for	1 [0-3]	1 [0-2]	1 [0-3]	
COVID-19 status <sup>c</sup> (range,				
days)				
RT-PCR+	255 (67%)	255 (88%)	0 (0%)	<0.001
RT-PCR-	126 (33%)	35 (12%)	91 (100%)	
Chest CT+	282 (74%)	282 (97%)	0 (0%)	< 0.001
Chest CT-	99 (26%)	8 (3%)	91 (100%)	

WDF+	264 (69%)	249 (86%)	15 (17%)	<0.001
WDF-	117 (31%)	41 (14%)	76 (84%)	
Lymphocyte count, 109/L				
Mean [range]	1.19 [0.08-4.90]	1.03 [0.08-4.22]	1.70 [0.14-4.90]	< 0.001
Median	1.01	0.96	1.05	
< 1.10 <sup>9</sup> /L, No. (%)	186 (49%)	159 (55%)	27 (30%)	< 0.001

Patient's baseline characteristics were compared by non-parametric tests, either the exact Fisher's test (qualitative) or the Kruskal-Wallis test (quantitative variables).

ICU: Intensive care unit; No.: Number of patients; NS: Not specified; WBC Differential Fluorescence scattergram (XN3100, SYSMEX $^{\$}$ )

<sup>&</sup>lt;sup>a</sup> Temporary unit in expectation of RT-PCR results (<24h);

<sup>&</sup>lt;sup>b</sup> Time interval since the onset of the first symptom.

<sup>&</sup>lt;sup>c</sup> Median time interval for COVID-19 status includes the completion time of RT-PCR, chest CT and CBC.